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CAMBRIDGE INTERNATIONAL EXAMINATIONS

Cambridge International Advanced Subsidiary and Advanced Level

MARK SCHEME for the May/June 2015 series

9700 BIOLOGY

9700/52

Paper 5 (Planning, Analysis and Evaluation), maximum raw mark 30

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Mark scheme abbreviations:

; separates marking points

I alternatives answers for the same point

R reject

A accept (for answers correctly cued by the question, or extra guidance)

AW alternative wording (where responses vary more than usual)

<u>underline</u> actual word given must be used by candidate (grammatical variants accepted)

max indicates the maximum number of marks that can be given

ora or reverse argument ecf error carried forward

I ignore

mp marking point (with relevant number)

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Qu	estion	Expected answer	Extra guidance	Mark
1	(a)	independent: concentration of amylase/enzyme; dependent: diameter / area of brown zone;	I amount R (amylase) extract A radius of brown zone A starch free/digested starch/clear zone	
			A area of brown zone minus well R blue zone	[2]
	(b)	any 8 from: independent variable 1 ref. to method of diluting the 0.5 g dm ⁻³ amylase/stock amylase solution and to give a minimum of 5 dilutions;	 0.0 and 0.5 g dm⁻³ can be included in the number of dilutions A serial/series/simple/proportional/dilution as method OR a description. Use the formula C₁V₁ = C₂V₂ to make A If the fungal extract is diluted instead of amylase but R mp5 	
		2 ref. to concentrations from 0.5 g dm ⁻³ downwards with correct units;	 minimum of 3 other stated values between 0.5 g dm⁻³ and 0.0 g dm⁻³ must correspond to dilution method chosen ecf if no method given A 10 fold or a 50% reduction serial dilution 	
		3 use of a <u>control</u> with example ; dependent variable	3 A water/0.0 g dm ⁻³ /boiled or denatured extract/enzyme	
		4 ref. to a suitable method of measuring diameters/width/radius/area of brown zones;	 4 e.g. using (suitable) ruler/callipers/string and ruler R metre ruler I graticules OR use a (transparent) grid/graph paper and count the number of squares OR take a photograph and measure using one the methods for finding area NOTE: method of measuring must match what they have stated they are measuring. 	

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5 ref. to testing the (fungal) extract/fungal amylase/fungal enzyme;	5	R if dilute the fungal extract	
6 plot a calibration curve of known concentrations and use it to determine extract concentration;	6	A compare extract result with results of known concentrations to identify extract concentration OR find approx. range of concentration and then do more at smaller intervals to identify extract	
standardising variables (max. 3)		concentration	
7 ref. to suitable stated volume/same volume of amylase (in each well);	7	If a volume is stated – max 1 cm ³ I amount / known	
8 leave (all plates) for same period of time;	8	if time stated, minimum 30 min/max 24 hours	
9 method of maintaining at same/constant/optimum/stated temperature;	9	e.g. incubator/constant temp. room A water bath	
		if temp. stated, any single temp. in range 15–65 °C I air conditioning	
10 use a buffer to keep the pH of the agar same ;	10	e.g. making or adding buffer to agar or starch (solutions) A adding buffer to the amylase solution before using it A if stated, any single pH	
11 same <u>concentration</u> of starch (in the agar plates);	11	I ref. to other nutrients in agar, I amount	
12 same depth/volume of agar in Petri dish;	12	A depth of agar plate	
13 cover to prevent contamination / evaporation ;		I mass of agar/depth of wells	
safety			
14 ref. to low risk investigation/hazard <u>and</u> suitable safety precaution;	14	fungal/enzyme allergy or fungus/enzyme/iodine/agar is irritant <u>and</u> wearing gloves/eye protection/mask I iodine as an allergen R no risk	
reliability			
15 ref. to a minimum of three replicates <u>and</u> calculate a mean or identify/eliminate/remove anomalies;	15	A original and 2 more/several/many/multiple A outliers for anomalies R reduce anomalies	[max 8]

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(c)	use a (glucose) biosensor/glucose dipstick or test-strip/named test strip, e.g. clinistix/uristix;	A use Benedict's test and EITHER weigh precipitate OR use colorimeter to find the intensity of blue solution left OR time the first appearance of a colour change A chromatography and stain (to show sugars) A Barfoed's test for monosaccharides	
	(because) $\gamma\text{-amylase}$ will produce glucose $\textbf{only}/\beta\text{-amylase}$ will produce maltose (mainly) ;	I idea that γ-amylase produces more glucose than β-amylase ${\bf R}$ ora	[2]
(d) (i)	1963 shown on extract B , plate 5 ;		
	1809 shown on extract D , plate 3;		[2]
(ii)	reject/eliminate/ignore/leave out (affected data from calculations);	A repeat until consistent results obtained I repeat unqualified OR repeat to find a mean	[1]

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(iii)	correct values for $\Sigma x = 1820$		
	and $\bar{x} = 303 / 303.3 / 303.3$	e.g. $\begin{array}{ c c c c c c }\hline \Sigma & 1820 & 342 \\\hline \hline \bar{x} & 303 & & & & & \\\hline \end{array}$	
	and $(x-\overline{x})^2$;		
	$(x-\overline{x})^2$ allowed 341 341.3 341.34 341.4 342		
	$(x-\bar{x})^2$ not allowed 341.32 341.2	allow ecf for s from wrong value of $\sum (x - \bar{x})^2$	
	correct value of s from table ;		
	correct values of s (±) 8.(0) 8.26 8.27 8.3		[max 2]
(iv)	any 3 from: 1 the larger/AW the brown area the more amylase/more enzyme activity ora	Ignore ref. to proportionality / positive correlation between enzyme concentration and gene copies	
	 OR the larger/AW the brown area the more gene copies; ora (person or extract) A had the highest/AW concentration of amylase/(person or extract) F has lowest AW concentration of amylase; (person) A has the highest number of gene copies/(person) F has the lowest number of gene copies; ref. to genetic variation in population (for the production of amylase); 	A strongest/largest/most/ ora weakest/smallest/least A A>B>C>D>E>F I copying results without any conclusion about rank order	
			[3]
		Total:	[20]

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2	(a)	any 3 from: variation in volunteers body mass/weight; ref. to use of drugs e.g. medication/self-inflicted; alcohol consumption; smoking status; ref. to ethnicity;	Mark the first three given. I environmental factors, e.g. temperature/light/noise in room/time of day I diet I ref. to distance between or position of electrodes/time of charge application	
		6 volunteer all have the same 'handedness';		
		7 no medical/named medical condition affecting nerve conduction ;	 e.g. Multiple Sclerosis(MS), Myalgic Encephalitis(ME), Muscular Dystrophy(MD), Motor Neurone Disease (MND),spinal bifida, polio A disease of the nervous system 	
		variation in method of applying test 8 same arm tested;	·	
		9 idea of volunteer not moving (during the test);	9 e.g. at rest/sitting/lying down	
		10 electrical charge/coulombs/voltage/potential difference;	10 A electrical stimulus I current	
		11 same number of volunteers in each age category;		[max 3]

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(b)	age/years	mean conduction velocity ± S _M	confidence limits		note: must have both numbers correct for one mark	
			lower limit	upper limit		
	60–69	52.2 ± 0.675	50.85	<u>53.55</u> ;		[1]
(c) (i)	30–39 and 70–79 OR 40–49 and 70–79 OR 50–59 and 70–79;				A descriptions, e.g. upper limit of 70–79 does not reach lower limit of 30–39 I error bars overlapping/range(bars) not overlapping I ref. to mean conduction velocity	
	there is no overlap of (confidence) limits/ S_M ;					[2]
(ii)	test: t test;				R continuous variation	
	reason: comparing (two) means/normal distribution/continuous data;					[2]
(iii)	there is no significant difference between the (mean) conduction velocities / NCV of (individuals from) different age groups;				needs to be clear that the significant difference is in the conduction velocity and the not the ages A the difference in (mean) conduction velocities of individuals from different age groups is not significant A stated age categories/women of different ages/young(er)	1
					and old(er) women/with age	1
(d)	large sample	size /number of peopl	e tested ;		A tested 394 individuals/many people/lots of people; I sufficient/enough people tested I number of different age categories/ref. to standard error	[1]
					Total:	[10]